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## Note

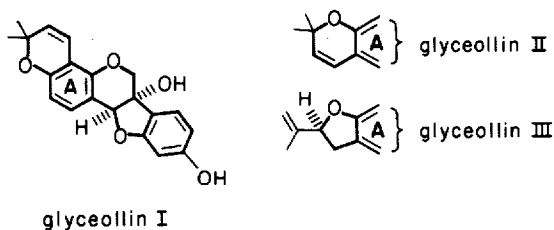
### Separation of glyceollin isomers I-III by thin-layer chromatography

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Environmental stress of soybeans by chemical treatment (herbicide<sup>1</sup>, heavy metal<sup>2</sup>, ozone<sup>3</sup>, etc.) or microbial infection<sup>2,4-15</sup> leads to accumulation of the anti-fungal metabolites glyceollin I, II and III (GI-III), the levels and relative ratios of which strongly depend on the plant tissue studied and the nature of the elicitor<sup>1,6,11,12</sup>. These isomeric phytoalexins can be analyzed by gas-liquid chromatography (GLC)<sup>4,8</sup> or high-performance liquid chromatography (HPLX)<sup>1,6,11,12,15</sup> but no separation is achieved with the usual thin-layer chromatographic (TLC) systems for phenolic compounds<sup>1-5,7-15</sup>. Accordingly, GI-III are usually quantitated after several TLC purification steps as "glyceollin", a mixture of the three compounds<sup>1-5,7-15</sup>.



The present study evaluated a large number of cellulose, polyamide or silica gel TLC systems suggested for analysis of phenols<sup>16,17</sup> but without success for separation of GI-III (data now shown). However, these compounds are easily separated by using multiple developments of formamide-impregnated silica gel layers, a sorbent described for TLC of *Umbelliferae* drugs<sup>18</sup>. Quantitation of GI-III in plant extracts is then achieved by using this system in place of the last purification-quantitation steps<sup>1-15</sup>, followed by normal UV spectrometry of the eluted spots. This method, when combined with radioautography, may also find application in radiotracer studies of the biosynthesis of GI-III.

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## EXPERIMENTAL

Cut soybean seedlings (5 g) were treated with the diphenyl ether herbicide acifluorfen (sodium 5-[2-chloro-4-(trifluoromethyl)phenoxy]-2-nitrobenzoate; 5 ppm, stem uptake for 72 h). The GI-III mixture was extracted with 40% ethanol and purified by TLC according to Ingham *et al.*<sup>11</sup>. The purified GI-III mixture and standards of GI, II and III (0.2, 1 and 5  $\mu\text{g}$ ; both separately and together) were spotted onto formamide-impregnated silica gel TLC plates [prepared by developing silica gel 60 F254, 0.25 mm thick TLC plates (Merck) twice with 5% formamide in acetone and drying at room temperature] and developed four times with diethyl ether-hexane (3:1). UV-absorbing spots of GI, II and III were detected at  $R_f$  values of 0.50, 0.42 and 0.35, respectively, and the compounds were quantitated by UV spectrophotometry (using the following absorption maxima and molar absorption coefficients: GI, 285 nm,  $8300 \text{ l mol}^{-1} \text{ cm}^{-1}$ ; GII, 285 nm,  $8700 \text{ l mol}^{-1} \text{ cm}^{-1}$ ; GIII, 292 nm,  $9600 \text{ l mol}^{-1} \text{ cm}^{-1}$ )<sup>4,6</sup> after scraping off the spots and eluting with  $2 \times 1 \text{ ml}$  ethanol. The levels of GI, II and III found in acifluorfen-treated soy bean seedlings were 7, 19 and 38  $\mu\text{g/g}$  fresh weight, respectively (compared to 4, 28 and 44  $\mu\text{g/g}$  fresh weight determined by HPLC in a separate experiment<sup>1</sup>). The GI-III content in the untreated leaves was  $< 1 \mu\text{g/g}$  fresh weight.

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## REFERENCES

- 1 T. K6mives and J. E. Casida, in preparation.
- 2 M. Yoshikawa, *Nature (London)*, 275 (1978) 546-547.
- 3 N. T. Keen and O. C. Taylor, *Plant Physiol.*, 55 (1975) 731-733.
- 4 N. T. Keen, J. J. Sims, D. C. Erwin, E. Rice and J. E. Partridge, *Phytopathology*, 61 (1971) 1084-1089.
- 5 J. J. Sims, N. T. Keen and V. K. Honwad, *Phytochemistry*, 11 (1972) 827-828.
- 6 R. L. Lyne, L. J. Mulheirn and D. P. Leworthy, *J. Chem. Soc., Chem. Commun.*, (1976) 497-498.
- 7 A. R. Ayers, J. Ebel, F. Finelli, N. Bergen and P. Albersheim, *Plant Physiol.*, 57 (1976) 751-759.
- 8 E. W. B. Ward, G. Lazarovits, C. H. Unwin and R. I. Buzzell, *Phytopathology*, 69 (1979) 951-955.
- 9 I. Paradies, J. R. Konze and E. E. Elstner, *Plant Physiol.*, 66 (1980) 1106-1109.
- 10 L. I. Weinstein, M. G. Hahn and P. Albersheim, *Plant Physiol.*, 68 (1981) 358-363.
- 11 J. L. Ingham, N. T. Keen, L. J. Mulheirn and R. L. Lyne, *Phytochemistry*, 20 (1981) 795-798.
- 12 P. Moesta and H. Grisebach, *Arch. Biochem. Biophys.*, 212 (1981) 462-467.
- 13 M. J. Holliday and N. T. Keen, *Phytopathology*, 72 (1982) 1470-1474.
- 14 G. Lazarovits and E. W. B. Ward, *Phytopathology*, 72 (1982) 1217-1221.
- 15 P. Moesta and H. Grisebach, *Physiol. Plant. Pathol.*, 21 (1982) 65-70.
- 16 R. Neher, in E. Stahl (Editor), *Thin Layer Chromatography*, Springer, New York, 1969, pp. 349-351.
- 17 K. Egger, in E. Stahl (Editor), *Thin Layer Chromatography*, Springer, New York, 1969, pp. 687-706.
- 18 T. Beyrich, *Planta Med.*, 13 (1965) 439-443.